Independent Project Week Supplies

**Student benches:**
- macropipettors, micropipettes, pipette tips, ep. tubes
- ep. tube holders
- turntable
- spreader & beaker, alcohol, fire extinguisher can
- alcohol lamp, matches
- test tube rack
- ice bucket
- parafilm/saran wrap/alum. foil

**Front/side benches:**
- blenders/large funnel/coffee filters
- knife/scalpels/forceps
- flasks/beakers/t.t. racks
- balances/weigh boats
- 70%/95% ETOH; 80% & 100% Acetone; Pet. Ether
- sterile toothpicks/swabs
- sterile discs
- latex gloves
- 50 ml/15 ml plastic cent. tubes
- plastic petri plates
- 13 X 100 test tubes, grad. cylinders
- 1 ml/5ml serol. pipettes
- stir plate, magnet (side of lab)
- fly morgues
- forceps, paint brushes, dissecting needles
- index cards
- mortars & pestles, sand
- slides/coverslips
- goggles
- chromatography tanks
- chromatography paper
- stopwatches
- ring stands with clamps, UV lamps
- lamps
- light box & included supplies for Hill reaction
- lux meters
- small funnels
- thermometers
- cellophane
- pH paper
**Incubators:** 5˚ C (refrig.)—both rooms; 13˚ C, 17˚ C (for sea urchins), and 37˚ C (for bacteria) incubators in A109
37 deg. shaker/incubator (containing overnight bacterial cultures) in A109
30 deg. shaker/incubator (for overnight yeast cultures) in A104
30 deg. constant temp. room—A108A—for yeast
25 deg. constant temp. room—A108B—for Drosophila

**Refrigerators:**

Supplies for peroxidase experiments:
3% H$_2$O$_2$—add 600 microliters to 20 ml. water to get working solution.
citrate buffer; TMB
DCIP
0.5M sucrose solution
stock solutions for making buffers of various pHs (see note on front desk for instructions)
blender bowls & tops
0.1 M NaOH and 0.1 M HCl
Sterile water (in tt racks)
rack of ep. tubes with 750 microl. of sterile water in each (FOR YEAST)

Also:
Euglena bottles on windowsill in A109
Plants & leaves from greenhouse/elsewhere: on table next to window in A109
Antibiotic solutions—in refrigerator in A111

**Chemicals et al.—on cart in A111:**

caffeine, nicotine, MSG, tobacco, ethanol, sodium benzoate, rubbing alcohol
confect. sugar, nutrasweet (Equal), saccharin (Sweet ’N Low); salt
Listerine, Scope, other mouthwashes
herbicide, insecticide
household cleaners—Formula 409, Lysol, bleach, Pine-sol
antibacterial first aid cream; antibacterial cleaning products, including toilet cleaner
nitric acid, sulfuric acid
sunscreens (various SPFs); tanning accelerator
antibacterial soap—several kinds; body washes, hand sanitizers; non anti-bact. soap
hair spray, bleach, ammonia, drain opener, vinegar
pepsin
Yellow #5, Blue #1 dyes; food coloring
Mannitol
Glucose, sucrose, fructose
Saline contact solutions
Yeast infection treatment
Vegetables, fruit et al.--A108C:
- turnip leaves, root
- kale
- red cabbage
- spinach
- garlic
- red leaf, iceberg, Romaine, Boston lettuce
- tea tree oil
- plain yogurt
- vanilla
- apple juice, lemon juice, cranberry juice, orange juice, lemonade
- mushroom
- apple jelly
- Coke, Pepsi, Mt. Dew, Mello Yellow, Blue Haw. Punch, yellow & blue Gatorade, yellow & blue Powerade
- carbonated water, Evian, 2% milk
- tuna,
- honeydew

Also in A108C: LB & NA plates (for bacteria); SC and SD plates (for yeast)

Useful pages in the Lab Manual:

- A-6 to A-11 Instructions for preparing written and oral reports on independent projects
- C-5 Extraction of plant pigments
- L113 web site Chromatography of ether-soluble plant pigments
- L113 web site Chromatography of water-soluble plant pigments
- C-12 Preparation of chloroplast suspension for Hill Reaction
- C-12 to C-13 Hill reaction
- L113 web site Preparation of crude peroxidase enzyme extract
- L113 web site Standard assay for peroxidase
- E-3 Simple Drosophila genetic map
- D-10 to D-12 Dilution, plating and UV irradiation of yeast
- L113 web site Lots of additional procedures provided